

Efficient Stereocontrolled Glycosidation of Secondary Sugar Hydroxyls by Silicon Tethered Intramolecular Glycosidation

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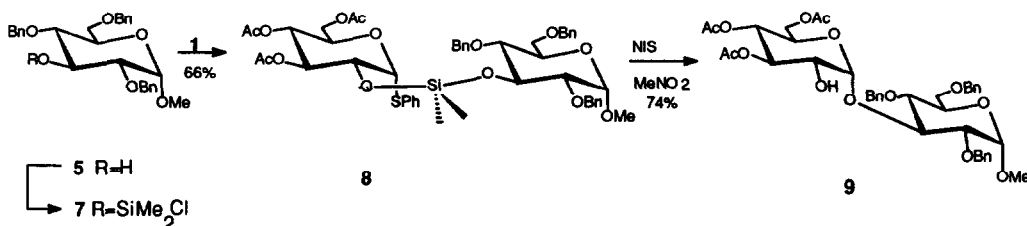
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Abstract: Disaccharides containing 1,2-*cis* glycoside linkages were synthesized by an efficient stereocontrolled two step process involving a silicon tethering step, to a dimethylsilyl acetal followed by intramolecular glycosidation with *N*-iodosuccinimide in nitromethane.

INTRODUCTION

The chemical synthesis of glycosides is of considerable potential interest in connection with many biological¹ and medical² studies, since glycosides occur widespread in nature. Therefore, a tremendous amount of literature on glycoside-synthesis exist,³ and especially during the last decade a number of very efficient methods of forming glycosidic linkages have been developed.⁴ The field is, however, still an area for specialists due to the high degree of unpredictability and lack of generality of the various methods. Contemporary intramolecular glycosidation methods,⁵⁻¹⁰ can be seen as an effort to achieve predictability through stereocontrol and enhanced reactivity. Recently intramolecular glycosidations have been achieved by carrying out the reaction with the aglycon tethered to the 2-position of the glycosyl-donor by a carbon^{5,6} or a silicon⁷⁻¹⁰ linkage. The tethering process is somewhat cumbersome, however, requiring 2 steps. In this paper a one step procedure for preparing dimethylsilylene tethered disaccharides from the hydroxy-sugars is described, as well as an improved method of intramolecular glycosidation allowing the efficient stereocontrolled glycosidation of secondary sugar alcohols.¹⁰

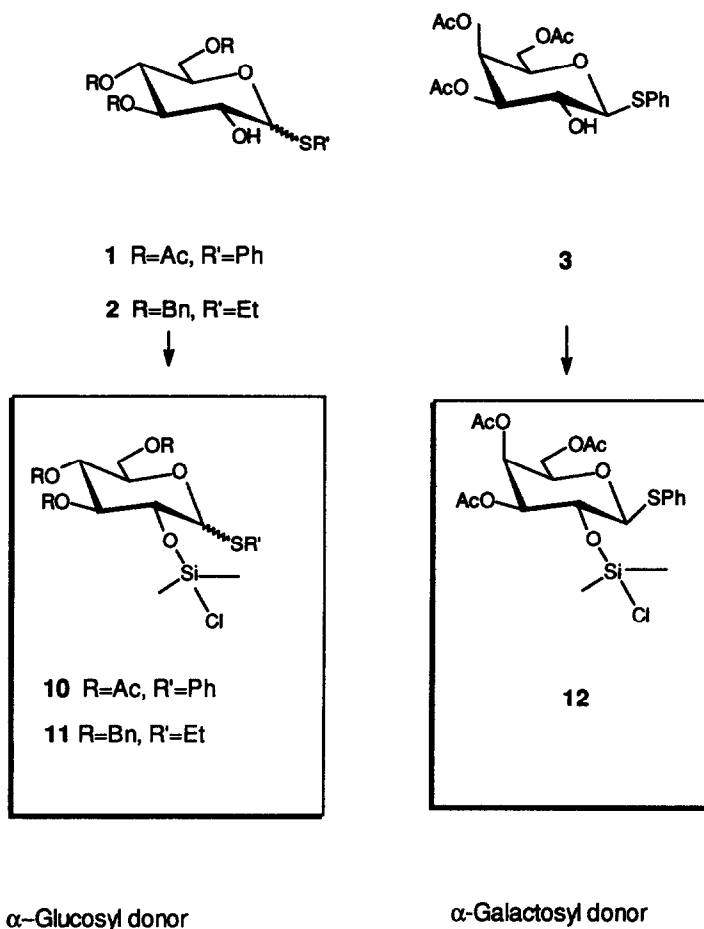
Scheme 1



RESULTS AND DISCUSSION

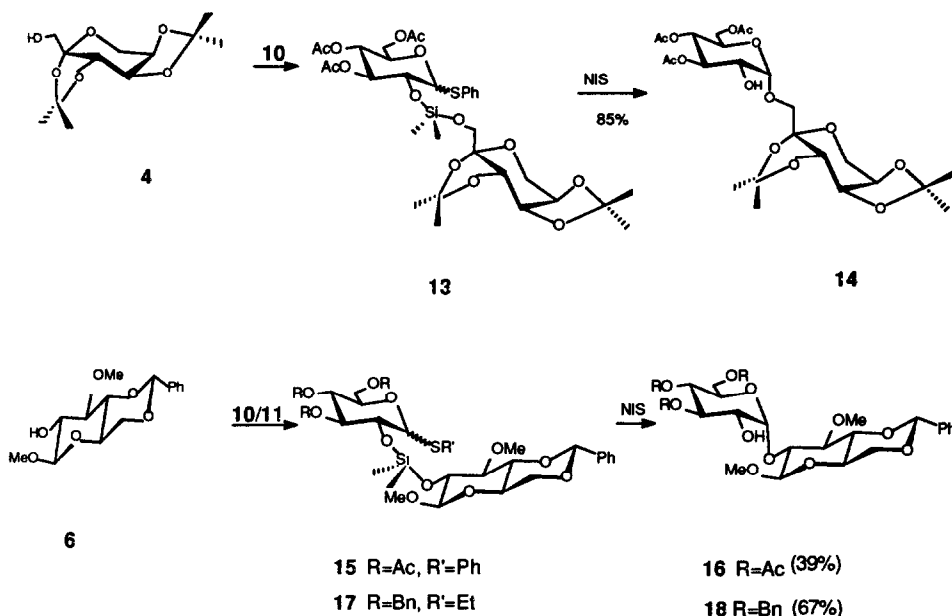
It has previously been described that aliphatic alcohols and phenols can be α -glucosidated using a thioglucoside, a dimethylsilylacetal linker with *N*-iodosuccinimide (NIS)/trifluoromethanesulfonic acid (TfOH) as promoter.^{8,9} As a continuation of this work it was decided to attempt to extend it to the synthesis of oligosaccharides. Thus, reaction of **5** (Scheme I) with excess dimethylsilyldichloride and triethylamine in dry ether allowed the isolation of the crude chlorodimethylsilyl ether **7**. The product contained several unidentified impurities, the formation of which it was not possible to avoid. Reaction of the thioglucoside **18,9** with 3.6 equiv of **7** gave the dimethylsilyl acetal **8** in 66% yield. If a smaller excess of **7** was employed the yield decreased probably due to complications caused by the impurities in **7**. The identity of the product **8** was easily recognised by the two downfield peaks (-1.4, -1.8) in the ¹³C NMR-spectrum from the diastereotopic methyl groups on the silylacetal.

Scheme II



Reaction of **8** with NIS and a catalytic amount of TfOH^{4b,4c} according to the previous protocol^{8,9} gave only 19% yield of α -glucoside **9**. In addition large amounts of **5** was isolated. Since NIS/TfOH had proved to be effective for the glycosidation of aliphatic alcohols regardless of steric hindrance, the low yield was probably not be caused by steric but by electronic effects, and could be due to the known reduced nucleophilicity of sugar hydroxy groups. Thus a slower intramolecular substitution could allow TfOH to cleave the silylene linkage faster than glycosidation could occur. If TfOH was omitted no activation of the thioglycoside occurred due to its "disarmed" nature. However by employing the dipolar, aprotic solvent MeNO₂ and increasing the temperature to 100°C **8** reacted with NIS alone to give the desired **9** in 74% yield. No β -glucoside was detected in the reaction product. At this point, improvement of the synthesis of the silylene linkage seemed in order. The most generally applicable synthesis would be to convert thioglycoside donors **1-3** into the chlorodimethylsilyl ethers **10-12** (Scheme II). Aglycons could then be converted to glycosides simply by facile silylation with **10, 11** or **12** followed by treatment with NIS. It was found that the unstable chlorodimethylsilyl ethers were handled much better when isolation was not attempted. Thus reaction

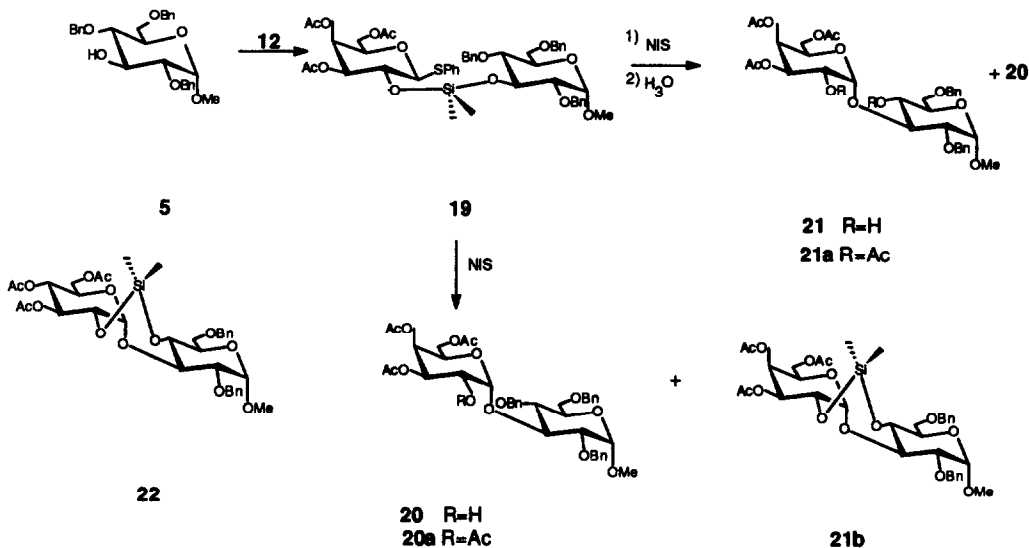
Scheme III



of **1** with 5 equiv. of Me₂SiCl₂ in pyridine/toluene followed by removal of excess Me₂SiCl₂ by distillation resulted in a solution of chlorosilyl ether **10** in pyridine/toluene. Alcohols could now easily be silylated with this solution (Scheme III). Thus reaction of the fructose-derivative **4**¹² with 1.5 equiv. of **10** gave the silyl acetal **13** in 72% yield, while the glucose-derivative **6**¹³ reacted with 1.6 equiv. of **10** to give silyl acetal **15** in 76% yield. Treatment of **13** with NIS in MeNO₂ at 100°C for 1 h gave the α -glucoside **14** in 85% yield. Reaction of **15**

with NIS in MeNO₂ at 100° was slow and required 4 h to run to completion giving 39% of α-glucoside **16**. In neither of the two cases were any β-glucoside detected. In the latter case the crude reaction product was, despite the low yield, extremely pure so the loss of material was probably caused by over-oxidation during the long reaction time. Therefore, the glycosidation was repeated with the armed glycosyl donor **2**, which could be reacted under milder conditions. A solution of chlorosilyl ether **11** was prepared from **2**, Me₂SiCl₂ and pyridine, and **6** was silylated with 1.25 equiv. of this solution resulting in a 93% yield of the silylacetal **17**. Reaction of **17** with NIS (2.5 equiv.) in MeNO₂ at 25°C for 4 h gave the α-glucoside **18** in 67% yield. The extension of the method to α-galactosidation was then attempted. Phenylthiogalactoside **3** was prepared in 56% yield from 3,4,6-tri-*O*-acetyl-β-D-galactosyl chloride¹⁴ by substitution with potassium benzenethiolate, and converted to a solution of the chlorodimethylsilyl ether **12** by reaction with excess

Scheme IV

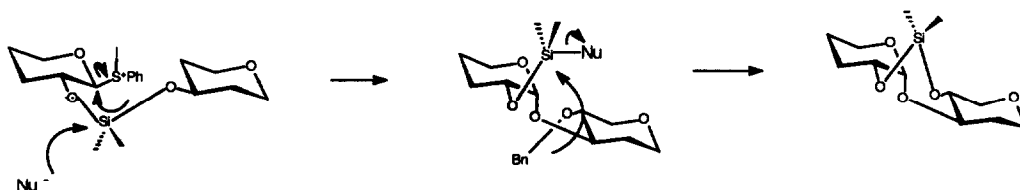


Me₂SiCl₂ and pyridine in toluene. Silylation of **5** with 1.5 equiv. of **12** gave the silylacetal **19** in 73% yield (Scheme IV). However, reaction of **19** with NIS (2.5 equiv.) in MeNO₂ for 1 2/3 h did not give one but two glycosides: the expected α-galactoside **20** and the 4-de-*O*-benzylated α-galactoside **21** in 32% and 49% yield, respectively. The structure of **21** was determined from the following facts: 1) It was an *O*-glycoside as seen from two ¹³C NMR signals at 100.7 and 97.9 ppm. 2) It was an α-galactoside as seen from the ¹H NMR doublet at δ 5.35 having a coupling constant of 3.5 Hz. 3) Acetylation of **20** and **21** introduced one acetyl group in **20** and two in **21** giving tetraacetate **20a** and pentaacetate **21a**, respectively. 4) The ¹H NMR spectrum of pentaacetate **21a** showed a double-doublet at low field (δ 5.05) with two large couplings (9.5 and 10.0 Hz) meaning that either OH-3 or OH-4 of **21** had been acetylated. 5) Irradiation of the triplet at δ 4.20 in the ¹H

NMR spectrum of **21a** converted the double doublets at 5.05 and 3.55 to doublets proving that OH-3 was the glycosylated hydroxy group.

The formation **21** was puzzling partly because no other debenzylated sugars were isolated and partly because debenzylation had not been observed in the reaction of **8** with NIS. Therefore the reaction of **19** was repeated, but the usual quenching with hydrochloric acid was avoided. In this case **20** and the tricyclic dimethylsilylene derivative **21b** was isolated in 31% and 46% yield respectively. Thus **21** was obviously formed from **21b** during work-up, and **21b** seemed the direct product of the debenzylation. To explain this, the mechanism outlined in Scheme V can be suggested. The intramolecular reaction must involve a nucleophilic substitution at silicon, probably by succinimide ($\text{Nu} = (\text{CH}_2\text{CO})_2\text{N}^-$), since the formation of a positive charge on silicon is unlikely. Reaction between the silylsuccinimide and I_2 formed in the reaction, could be expected to lead to NIS and a silyl iodide known to be a debenzylation agent. Finally intramolecular debenzylation would lead to **21b**. Apparently the formation of the 8 membered ring was not unfavorable.

Scheme V



As a result of this mechanism, debenzylation should be a process occurring after the glycosidation. To investigate this, the reaction of **8** with NIS was reinvestigated. Reaction for 3 h in MeNO_2 at 100°C , followed by non-acidic work-up led to α -glucoside **9** (29%) and 4,3'-dimethylsilylene derivative **22** (22%, Scheme IV). So the debenzylation appeared to be general, but much slower in this case.

In conclusion, this paper describes an efficient protocol for stereocontrolled synthesis of α -glucosides or α -galactosides in oligosaccharides by a two step intramolecular procedure: An improved method of preparing dimethylsilyl linked disaccharides allows easy synthesis of the precursors for intramolecular glycosidation which is efficiently carried out with NIS in MeNO_2 . In difficult cases use of an "armed" glycosyl donor improves the yield.

EXPERIMENTAL

^{13}C -NMR spectra were recorded on a Bruker AC-250 instrument with D_2O as solvent using 1,4-dioxane as the internal reference (67.40 ppm). Melting points are uncorrected. Optical rotations were measured on a Perkin Elmer 141 polarimeter. Microanalyses were carried out by Leo Microanalytical Laboratory. Concentrations were performed on a rotary evaporator at a temperature below 40°C .

Ethylthio 3,4,6-tri-*O*-benzyl- β -D-glucopyranoside (2). Ethylthio 2-*O*-acetyl-3,4,6-tri-*O*-benzyl- β -D-glucopyranoside (3.1 g, 5.8 mmol), prepared from 3,4,6-tri-*O*-benzyl-1,2-*O*-(methoxyethylidene)- α -D-glucopyranose¹⁵ using the analogous procedure described for mannose¹⁶, was dissolved in dry MeOH (40 ml), and Na (50 mg) in 10 ml MeOH was added. After stirring for 18 h the solution was neutralized by addition of excess Amberlite IR-120 cation-exchange resin (H^+), stirring until neutral, filtration and concentration to

give sirupy **2** (2.71 g, 95%). Crystallization from ether-pentane gave 2.07 g (72%). Mp: 61-62°C [α]_D²⁰ -14.4° (c 1.0, CH₂Cl₂) ¹³C NMR (CDCl₃): δ 137.9-138.4, 127.4-128.3 (Ar's), 85.9 (C-1), 85.9, 79.3, 77.3, 75.1, 74.9, 73.3, 73.1 (C-2-C-5, Bn's), 68.9 (C-6), 24.1 (CH₂S), 15.3 (Me). ¹H NMR (CDCl₃): δ 7.1-7.4 (m, 15H, Ar's), 4.8-5.0 (m, 3H, Bn's), 4.5-4.6 (m, 3H, Bn's), 4.3 (d, 1H, J₁₂ = 9 Hz, H-1), 3.4-3.8 (m, 6H, H-2-H-6b), 2.70 (2 dq, 2H, CH₂S), 1.30 (t, 3H, J = 7.0 Hz, Me). Anal. Calc for C₂₉H₃₄O₅S: C, 70.42; H, 6.93. Found: C, 70.43; H, 6.85.

Phenylthio 3,4,6-tri-O-acetyl- β -D-galactopyranoside (3). A solution of K (153 mg, 3.9 mmol) in dry MeOH (5 ml) was prepared under Ar, and cooled to -78°C. Thiophenol (0.55 g, 5 mmol, 1.3 eq) was added, and after 10 min at this temperature 3,4,6-tri-O-acetyl- β -D-galactopyranosyl chloride¹⁴ (1.16 g, 3.6 mmol) in MeOH (3 ml) was added. The mixture was allowed to reach 25°C, and then stirred for 6 h. Concentration, addition of EtOAc (50 ml), washing of the organic layer with NaHCO₃-solution (25 ml), drying and concentration left a sirup (1.37 g). Flash-chromatography (EtOAc-Pentane 1:2) gave **3** (803 mg, 56 %) as a faster moving product. Crystallization from ether gave mp. 117-119°C. [α]_D²⁰ + 10.7° (c 1.3, CH₂Cl₂). ¹³C NMR (CDCl₃): δ 132.8, 128.9, 128.3 (Ar), 88.7 (C-1), 74.4, 73.7, 67.3, 66.9 (C-2, C-3, C-4, C-5), 61.7 (C-6), 20.4-6 (3C, Ac's). ¹H NMR (CDCl₃) δ 7.6 (m, 2H, Ar), 7.3 (m, 3H, Ar), 5.4 (dd, 1H, J₃₄ = 4.0 Hz, J₄₅ = 1.0 Hz, H-4), 4.95 (dd, 1H, J₂₃ = 9.5 Hz, H-3), 4.65 (d, 1H, J₁₂ = 9.5 Hz, H-1), 4.2 (dd, 1H, J_{66'} = 11.5 Hz, J₅₆ = 7.0 Hz, H-6), 4.1 (dd, 1H, J_{56'} = 6.0 Hz, H-6'), 3.95 (ddd, 1H, H-5), 3.8 (t, 1H, H-2), 2.5 (bs, 1H, OH), 2.0-2.1 (3s, 9H, Ac's). Anal. Calc. for C₁₈H₂₂O₈S: C, 54.26; H, 5.57. Found: C, 53.97; H, 5.68. Eluting the column with EtOAc gave a slower moving product of methyl 3,4,6-tri-O-benzyl- α , β -D-glucopyranoside (246 mg).

Methyl 3-O-chlorodimethylsilyl-2,4,6-tri-O-benzyl- α -D-glucopyranoside (7). To methyl 2,4,6-tri-O-benzyl- α -D-glucopyranoside¹¹ (**5**, 4.64 g, 10 mmol) in dry ether (5 ml) under N₂ at 0°C was added dimethylsilyldichloride (2.41 ml, 258 g, 20 mmol) followed by Et₃N (1.39 ml, 1.01 g, 10 mmol). After stirring 10 min at 0°C, filtration and concentration of the filtrate left a sirupy residue of crude **7** (5.19 g, 93%). ¹³C NMR (CDCl₃): δ 137.9-138.5 and 127.2-128.4 (Ar), 97.7 (C-1), 79.3, 77.9, 75.6, 74.7, 73.4, 73.3, 69.5 (C-2, C-3, C-4, C-5, 3 Bn), 54.8 (OMe), 2.9, 2.7 (Me₂Si).

Phenylthio 2-O-dimethyl-(1-O-methyl-2,4,6-tri-O-benzyl- α -D-glucopyranos-3-oxyl)-silyl-3,4,6-tri-O-acetyl- α -D-glucopyranoside (8). Phenylthio 3,4,6-tri-O-acetyl- α -D-glucopyranoside⁹ (**1**, 100 mg, 0.25 mmol) dissolved in pyridine (0.5 ml) and THF (2 ml) was stirred under N₂. A solution of **7** (502 mg, 0.9 mmol, 3.6 eq.) in a small amount of THF (0.2 ml) was added. After stirring 2 h at 25°C ether (50 ml) was added, and the solution washed with water (10 ml) and 10% NaCl-solution (10 ml). Drying (MgSO₂), filtration and concentration gave a sirupy residue (553 mg). Flash-chromatography in EtOAc-pentane 1:10 followed by 1:6 followed by 1:4 gave the silyl acetal **8** (151 mg, 66%). [α]_D²⁰ + 137.0° (c 0.4, CH₂Cl₂). ¹³C NMR (CDCl₃): δ 170.3, 169.7, 169.7 (Ac's), 137.6-138.1 (Ar), 126.8-131.0 (Ar), 97.8 (C-1), 87.9 (C-1'), 79.7, 78.3, 74.6, 74.2, 73.3 (2C), 73.1, 70.3, 69.5, 68.6, 68.4, 67.9 (C-2 - C-6, C-2'-C-5', Bn's), 62.1 (C-6'), 54.8 (OMe), 20.5-20.7 (Ac's), -1.4, -1.8 (Me₂Si). ¹H NMR (CDCl₃): δ 7.2-7.5 (m, 20 H, Ar's), 5.8 (d, 1H, J_{1'2'} = 5.5 Hz, H-1'), 5.3 (t, 1H, J_{3'4'} = J_{4'5'} = 9.5 Hz, H-4'), 4.95 (dd, 1H, J_{2'3'} = 10 Hz, H-3'), 4.90 (d, 1H, Bn), 4.80 (d, 1H, Bn), 4.1-

4.7 (m, 9H), 4.0 (dd, 1H, $J_{6'a6'b} = 12.0$ Hz, $J_{5'6'a} = 2.0$ Hz, H-6a'), 3.3-3.7 (m, 4H), 3.35 (dd, 1H, H-2'), 3.3 (s, 3H, MeO), 2.0-2.1 (3s, 9H, Ac's), 0.25, 0.20 (2s, 6H, Me₂Si).

Anal. Calc. for C₄₈H₅₈O₁₄Si x H₂O: C, 61.52; H, 6.45. Found: C, 61.69; H, 6.37.

Methyl 3-O-(3,4,6-tri-O-acetyl- α -D-glucopyranosyl)-2,4,6-tri-O-benzyl- α -D-glucopyranoside (9). To a solution of **8** (93 mg, 0.10 mmol) in MeNO₂ (5 ml) was added N-iodosuccinimide (75 mg, 3.3 eq), and the mixture was refluxed 1 h. Hydrochloric acid (25 ml, 1M) was added, and the water-layer was extracted with EtOAc (3 x 25 ml). The combined organic layers were washed with NaHCO₃ (10 ml, saturated) and Na₂S₂O₃-solutions (10 ml, 5%). Drying (Na₂SO₄) filtration and concentration gave a syrup (102 mg). Flash-chromatography in EtOAc-Pentane 1:4 followed by 1:2 followed by 1:1 gave the glycoside **9** (56 mg, 74%) as a clear syrup. $[\alpha]_D^{20} + 9.8^\circ$ (c 1.1, CH₂Cl₂), ¹³C NMR (CDCl₃): δ 171.1, 170.7, 169.5 (Ac's), 137.5, 127.6-128.5 (Ar), 98.4 (C-1'), 97.3 (C-1), 78.6, 77.9, 76.6, 74.1, 73.6, 73.5, 72.4, 71.1, 69.8, 68.1, 67.7, 67.5 (C-2 - C-6, C-2' - C-5', Bn's), 61.5 (C-6'), 55.0 (OMe), 20.6-20.8 (Ac's), ¹H NMR (CDCl₃): δ 7.1-7.4 (m, 15H, Ar's), 5.45 (d, 1H, $J_{1'2'} = 3.5$ Hz, H-1'), 5.25 (t, 1H, $J_{3'4'} = J_{4'5'} = 9.5$ Hz, H-4'), 4.95 (t, 1H, $J_{2'3} = 10$ Hz, H-3'), 4.8 (d, 1H, Bn), 4.75 (d, 1H, $J_{12} = 3.5$ Hz, H-1), 4.7 (d, 1H, Bn), 4.4 - 4.6 (m, 5H), 4.15 (m, 1H), 3.6 - 3.9 (m, 7H), 3.5 (dd, 1H, H-2'), 3.35 (s, 3H, OMe), 2.0-2.1 (3s, 9H, Ac's).

Anal. Calc. for C₄₀H₄₈O₁₄ x H₂O: C, 62.33; H, 6.54. Found: C, 62.23; H, 6.38.

Reagent 10 (" α -Glu"). To thioglucoside **1⁹** (2.33 g, 5.9 mmol) in dry toluene (40 ml) and dry pyridine (10 ml) under Ar was added dimethylsilyldichloride (3.5 ml, 3.78 g, 29 mmol, 5 eq), and the mixture was stirred for 1 h at 25°C. The apparatus was arranged for distillation and 10 ml was distilled off at which time the temperature at the top of the column had reached 105°C. The solution was cooled to 25°C and used as reagent **10** (40 ml, 0.15 M).

Reagent 11 ("armed α -Glu") To thioglucoside **2** (1.28 g, 2.6 mmol) in dry pyridine (5 ml) and dry toluene (20 ml) under Ar was added dimethylsilyldichloride (1.6 ml, 1.68 g, 13 mmol, 5 eq). After stirring 1 h at 25°C, the mixture was distilled carefully until the temperature reached 105-106°C (ca 15 ml). The solution in the distillation flask was cooled to 25°C and used as reagent **11** (11.6 ml, 0.22M).

Reagent 12 (" α -Gal"). To thiogalactoside **3** (719 mg, 1.8 mmol) in dry pyridine (3.5 ml) and dry toluene (15 ml) under Ar was added dimethylsilyldichloride (1.1 ml, 1.18 g, 9 mmol, 5 eq), and the mixture was stirred at 25°C for 1 h. Careful distillation until the temperature in the column reached 108°C (ca. 10 ml distilled) followed by cooling left a solution of reagent **12** (10 ml, 0.18 M).

Phenylthio 2-O-(2,3:4,5-di-O-isopropylidene- β -D-fructopyranos-1-oxo)dimethylsilyl-3,4,6-tri-O-acetyl- α , β -D-glucopyranoside (13). 2,3:4,5-di-O-isopropylidene- β -D-fructopyranose¹² (**4**, 0.50 g, 1.92 mmol) in pyridine (5 ml) was treated with reagent **10** (2.9 mmol **10** in 20 ml, 1.5 eq) and stirred under Ar at 25°C for 18 h. Ether (100 ml) was added, and the solution washed with H₂O (20 ml) and NaCl-solution (10%, 20 ml). The water layers were extracted with ether (10 ml), and the combined organic layers were dried (Na₂SO₂), filtered and concentrated to a syrup (1.99 g). Flash-chromatography in EtOAc-pentane 1:4 followed by 1:3 gave pure **13** (0.982 g, 72%, α/β 1:4). $[\alpha]_D^{20} + 9.5^\circ$ (c 2.4, CH₂Cl₂). ¹³C NMR (CDCl₃): 169.6-170.5 (Ac's), 127.7-

132.8 (Ar), 108.8, 108.2 (Me₂C's), 102.9 (C-2), 88.6 (C-1'), 75.4, 71.6, 71.0, 70.1, 69.2, 68.5 (6 C), 63.8, 62.3, 61.0 (C-1, C-6, C-6'), 26.5, 25.8, 25.5, 23.9 (Me₂C's), 20.5-20.9 (Ac's), -2.6, -2.9 (Me₂Si). ¹H NMR (CDCl₃): δ 7.5 (m, 2H, Ar), 7.3 (m, 3H, Ar), 5.1 (t, 1H, J_{3'4'} = J_{4'5'} = 9.5 Hz, H-4'), 4.95 (t, 1H, J_{1'2'} = 9.5 Hz, H-3'), 4.65 (d, 1H, J_{1'2'} = 9.5 Hz, H-1'), 4.55 (dd, 1H, J₂₃ = 8 Hz, J₃₄ = 2.5 Hz, H-4), 4.4 (d, 1H, H-3), 3.7-4.4 (m, 9H), 2.0-2.1 (3s, 9H, Ac's), 1.3-1.6 (4s, 12H), 0.25, 0.15 (2s, 6H, Me₂Si).
Anal. Calc. for C₃₂H₄₆O₁₄SiS x H₂O: C, 52.44; H, 6.60. Found: C, 52.46; H, 6.41.

2,3:4,5-di-O-isopropylidene-1-O-(3,4,6-tri-O-acetyl-α-D-glucopyranosyl)-β-D-fructopyranoside (14).

To a solution of **13** (345 mg, 0.48 mmol) in MeNO₂ (15 ml) was added N-iodosuccinimide (272 mg, 1.2 mmol, 2.5 eq), and the mixture was refluxed for 1 h. Hydrochloric acid (1M, 50 ml) was added, and the mixture was extracted with EtOAc (5 x 25 ml). The combined organic layers were washed with saturated NaHCO₃-solution (10 ml) and Na₂S₂O₃-solution (10%, 10 ml), dried (MgSO₄) and concentrated to a syrup (394 mg). Flash-chromatography in EtOAc-pentane 1:2 followed by 1:1 gave **14** as a clear syrup (224 mg, 85%). [α]_D²⁰ + 66.3° (c 2.7; CH₂Cl₂). ¹³C NMR (CDCl₃): δ 171.0, 170.5, 169.5 (Ac's), 109.0, 108.8 (Me₂C's), 101.7 (C-2), 98.1 (C-1'), 73.2, 70.8, 70.7, 70.0, 69.9, 69.1, 67.8, 67.7 (C-1, C-3 - C-5, C-1' - C-5'), 61.7, 61.1 (C-6, C-6'), 26.4, 25.8, 25.5, 23.9 (Me₂C's), 20.7, 20.6, 20.5 (Ac's), ¹H NMR (CDCl₃): δ 5.25 (t, 1H, J_{3'4'} = J_{4'5'} = 9.5 Hz, H-4'), 5.05 (t, 1H, J_{2'3'} = 9.5 Hz, H-3'), 5.00 (d, 1H, J_{1'2'} = 3.5 Hz, H-1'), 4.60 (dd, 1H, J₄₅ = 8.0 Hz, J₃₄ = 2.5 Hz, H-4), 4.40 (d, 1H, H-3), 4.25 (m, 2H), 4.10 (m, 2H), 3.95 (dd, 1H, J_{6a6b} = 13.0 Hz, J_{56a} = 2.0 Hz, H-6a), 3.90 (d, 1H, J_{1a1b} = 10.5 Hz, H-1a), 3.75 (dd, 1H, J_{56b} = 0.5 Hz, H-6b), 3.70 (dd, 1H, H-2'), 3.55 (d, 1H, H-1b), 2.0-2.1 (3s, 9H, Ac's), 1.3-1.55 (4s, 12 H, Me₂C's).
Anal. Calc. for C₂₄H₃₆O₁₄ x 2 H₂O: C, 49.31; H, 6.90. Found: C, 49.57; H, 6.53.

Phenylthio 2-O-(4,6-O-benzylidene-1,3-di-O-methyl-β-D-glucopyranos-2-oxo)dimethylsilyl-3,4,6-tri-O-acetyl-α,β-D-glucopyranoside (15). A solution of methyl 4,6-O-benzylidene-3-O-methyl-β-D-glucopyranoside **13** (6, 0.52 g, 1.76 mmol) in dry pyridine (5 ml) under Ar was treated with reagent **10** (20 ml, 2.9 mmol, 1.65 eq), and stirred for 18 h. Ether (100 ml) was added, and the organic layer washed with water (20 ml) and NaCl-solution (10%, 20 ml). The combined aqueous layers were extracted with ether (10 ml), and the combined organic layers dried (Na₂SO₄) and concentrated to a syrup (2.01 g). Flash-chromatography in EtOAc-pentane 1:4 followed by 1:3 gave pure **15** (1.00 g, 76%, α/β 1:3) as a clear syrup. [α]_D²⁰ + 6.7 (c 1.4; CH₂Cl₂). ¹³C NMR (CDCl₃): δ 171.0, 170.0, 169.6 (Ac's), 125.9-132.0 (Ar's), 104.6 (Ph-CH), 101.0 (C-1), 88.8 (C-1'), 82.8, 81.7, 75.4, 75.2, 71.0, 68.7, 65.8 (7 C), 62.4, 60.9, 57.2 (C-6', MeO's), 20.5-21.1 (Ac's), -1.6, -1.7 (Me₂Si). ¹H NMR (CDCl₃): δ 7.2-7.6 (m, 10H, Ar's), 5.55 (s, 1H, PhCH), 5.1 (t, 1H, J_{3'4'} = J_{4'5'} = 9 Hz, H-4'), 4.95 (t, 1H, J_{2'3'} = 9.5 Hz, H-3'), 4.65 (d, 1H, J_{1'2'} = 9.5 Hz, H-1'), 3.3-4.4 (m, 17 H), 2.0-2.1 (3s, 9H, Ac's), 0.2 (2s, 6H, Me₂Si). MS (CI, NH₃): m/z 768 (M + NH₄⁺).

Methyl 4,6-O-benzylidene-3-O-methyl-2-O-(3,4,6-tri-O-acetyl-α-D-glucopyranosyl)-β-D-glucopyranoside (16). To a solution of **15** (110 mg, 0.15 mmol) in MeNO₂ (5 ml) was added N-iodosuccinimide (83 mg, 0.37 mmol, 2.5 eq), and the mixture was refluxed for 4 h. Hydrochloric acid (1M, 25 ml) was added, and the mixture was extracted with EtOAc (3 x 25 ml). The combined organic layers were washed with NaHCO₃ (saturated, 10 ml) and Na₂S₂O₃ (10%, 10 ml). Drying (MgSO₄) and concentration left a syrupy residue, that was purified by Flash-chromatography in EtOAc-pentane 1:2 followed by 1:1 to give **16**

as a syrup (33 mg, 39%). $[\alpha]_D^{20} + 65.6^\circ$ (c 1.1, CH_2Cl_2), ^{13}C NMR (CDCl_3): 169.5-171.0 (Ac's), 136.9, 125.8-129.5 (Ar), 104.0 (PhCH), 101.0 (C-1), 98.8 (C-1'), 81.6, 81.1, 78.6, 73.2, 70.8, 68.4, 67.8, 67.6, 65.7 (C-2 - C-6, C-2' -C-5'), 61.6, 60.5, 57.2 (C-6', OMe's), 20.5-20.7 (Ac's), ^1H NMR (CDCl_3): δ 7.4-7.5 (m, 5H, Ar), 5.55 (s, 1H, PhCH), 5.30 (d, 1H, $J_{1'2'} = 4.0$ Hz, H-1'), 5.25 (t, 1H, $J_{3'4'} = J_{4'5'} = 9.5$ Hz, H-4'), 5.05 (t, 1H, $J_{2'3'} = 9.5$ Hz, H-3'), 4.45 (d, 1H, $J_{12} = 7.5$ Hz, H-1), 4.2-4.4 (m, 3H), 4.1 (m, 1H), 3.3-3.9 (m, 6H), 3.6, 3.55 (2s, 6H, OMe's), 2.0 (3s, 9H, Ac's). MS (CI, NH_3): m/z 602 (M + NH_4^+).

Ethylthio 2-O-(4,6-O-benzylidene-1,3-di-O-methyl- β -D-glucopyranos-2-oxo)dimethylsilyl-3,4,6-tri-O-benzyl- β -D-glucopyranoside (17). To a solution of methyl 4,6-O-benzylidene-3-O-methyl- β -D-glucopyranoside¹³ (0.40 g, 1.35 mmol) in pyridine (1 ml) was added reagent **11** (7.6 ml, 0.22M, 1.7 mmol, 1.25 eq) the mixture was stirred for 18 h at 25°C under Ar. Ether (50 ml) was added, and the solution was washed with water (10 ml) and NaCl-solution (10 ml, 10%). Drying (Na_2SO_2) and concentration left a syrup (1.52 g). Flash-chromatography in EtOAc-pentane 1:10 followed by 1:6 followed by 1:4 gave **17** as a colorless syrup (1.06 g, 93%). $[\alpha]_D^{20} -33.8^\circ$ (c 4.5, CH_2Cl_2). ^{13}C NMR (CDCl_3): δ 137.0 - 138.7, 125.6 - 128.4 (Ar), 104.2 (PhCH), 100.5 (C-1), 86.7, 85.8, 85.7, 82.7, 81.0, 78.7, 74.7, 74.4, 73.6, 72.8, 68.5, 68.2, 65.2 (13 C), 60.5, 56.7 (OMe's), 24.1, 14.7 (SEt). Anal. Calc. for $\text{C}_{46}\text{H}_{58}\text{O}_{11}\text{SiS} \times \text{H}_2\text{O}$: C, 63.86; H, 6.99. Found: C, 63.81; H, 6.87.

Methyl 4,6-O-benzylidene-3-O-methyl-2-O-(3,4,6-tri-O-benzyl- α -D-glucopyranosyl)- β -D-glucopyranoside (18). To a solution of **17** (583 mg, 0.69 mmol) in MeNO_2 (20 ml) under Ar was added N-iodosuccinimide (387 mg, 1.72 mmol, 2.5 eq), and the mixture was stirred for 4 h. Hydrochloric acid (1 ml, 1M) was added, and after 10 min. stirring CH_2Cl_2 (200 ml) was added. The organic layer was washed with HCl (50 ml, 1M), NaHCO_3 -solution (50 ml, saturated) and $\text{Na}_2\text{S}_2\text{O}_3$ -solution (50 ml, 10%), dried (MgSO_4) and concentrated. Flash-chromatography in EtOAc-pentane 1:4 followed by 1:2 gave **18** as a colorless syrup (337 mg, 67%). $[\alpha]_D^{20} + 40.1^\circ$ (c 0.09, CH_2Cl_2). ^{13}C NMR (CDCl_3): δ 138.0-138.7, 125.9-128.9, 104.4 (PhCH), 101.1 (C-1), 99.5 (C-1'), 83.2, 81.8, 81.4, 78.7, 77.2, 75.1, 74.8, 73.4, 73.2, 70.9, 68.6, 68.3, 65.9 (C-2 - C-6, C-2'-C-6', Bn's), 60.8, 57.2 (OMe's). ^1H NMR (CDCl_3): δ 7.1-7.6 (m, 20H, Ar), 5.55 (s, 1H, PhCH), 5.25 (bs, 1H, H-1'), 4.3-5.0 (m, 10H), 4.05 (m, 1H), 3.3-3.9 (m, 8H), 3.55, 3.50 (2s, 6H, OMe's). MS (CI, NH_3): m/z 746 (M + NH_4^+).

Phenylthio 2-O-dimethyl-(1-O-methyl-2,4,6-tri-O-benzyl- α -D-glucopyranos-3-oxo)silyl-3,4,6-tri-O-acetyl- β -D-galactopyranoside (19). To a solution of methyl 2,4,6-tri-O-benzyl- α -D-glucopyranoside (559 mg, 1.2 mmol) in THF (2 ml) under Ar was added reagent **12** (10 ml, 1.8 mmol, 1.5 eq), and the mixture was stirred at 25°C for 18 h under Ar. Ether (100 ml) was added, and the solution was washed with water (20 ml) and NaCl-solution (20 ml, 10%). The combined aqueous layers were extracted with ether (10 ml), and the organic layers were dried (Na_2SO_4) and concentrated to a syrup (1.57 g). Flash chromatography in EtOAc-pentane 1:10 followed by 1:4 followed by 1:3 gave **19** as a colorless syrup (804 mg, 73%). $[\alpha]_D^{20} + 33.0^\circ$ (c 1.9, CH_2Cl_2). ^{13}C NMR (CDCl_3): δ 170.0, 127.3-133.4 (Ar's), 97.8 (C-1), 88.6 (C-1'), 79.6, 78.1, 74.7, 74.4, 73.8, 73.3, 72.9, 69.5, 68.4, 68.2, 67.4, (11C), 61.6 (C-6'), 54.8 (OMe), 20.5-20.8 (Ac's), -1.7, -2.3 (Me_2Si). ^1H NMR (CDCl_3): δ 7.1-7.5 (m, 20H, Ar's), 5.35 (dd, 1H, $J_{3'4'} = 3.0$ Hz, $J_{4'5'} = 0.5$ Hz, H-4'), 4.90 (d, 1H, $J = 11.0$ Hz, Bn), 4.75 (d, 1H, $J_{1'2'} = 9.5$ Hz, H-1'), 4.70 (d, 1H, $J_{12} = 3.5$ Hz, H-1). 4.3-4.7 (m, 5H), 4.0-4.2

(m, 5H), 3.90 (t, 1H, $J = 9.5$ Hz), 3.5-3.7 (m, 4H), 3.45 (dd, 1H, $J_{23} = 9.5$ Hz, H-2), 3.30 (s, 3H, OMe), 2.05-2.1 (3s, 9H, Ac's), 0.3, 0.1 (Me₂Si). Anal. Calc. for C₄₈H₅₈O₁₄SiS x H₂O: C, 61.52; H, 6.45. Found: C, 61.52; H, 6.35.

Methyl 3-O-(3,4,6-tri-O-acetyl- α -D-galactopyranosyl)-2,4,6-tri-O-benzyl- α -D-glucopyranoside (20) and methyl 2,6-di-O-benzyl-3-O-(3,4,6-tri-O-acetyl- α -D-galactopyranosyl)- α -D-glucopyranoside (21). To a solution of **19** (185 mg, 0.20 mmol) in MeNO₂ (5 ml) under Ar was added N-iodosuccinimide (113 mg, 0.50 mmol, 2.5 eq). The mixture was refluxed for 1 h and 40 min., and after cooling to 25°C aqueous HCl (1 ml, 1M) was added. After stirring 10 min NaHCO₃-solution (25 ml, saturated) was added, and the mixture was extracted with EtOAc (3 x 25 ml). The combined organic layers were washed with Na₂S₂O₃-solution (10 ml, 10%), dried (MgSO₄) and concentrated to give a clear syrup (183 mg). Flash-chromatography in EtOAc-pentane 1:4 followed by 1:3 followed by 1:2 gave as a colorless syrup **20** (48 mg, 32%). Eluting with EtOAc gave **21** (66 mg, 49%) as a colorless syrup. Total yield: 81%. **20**: [α]_D²⁰ + 106.4° (c 2.3, CH₂Cl₂), ¹³C NMR (CDCl₃): δ 170.6, 170.4, 170.1 (Ac's), 137.6, 137.4, 127.6-128.3 (Ar), 98.9 (C-1'), 97.4 (C-1), 78.5, 78.2, 76.6, 74.2, 73.6, 72.7, 70.6, 69.7, 68.4, 68.0, 67.4, 66.7 (C-2 - C-6, C-2' - C-5', Bn's), 61.6 (C-6'), 55.0 (OMe), 20.5-20.7 (Ac's). ¹H NMR (CDCl₃): δ 7.1-7.4 (m, 15 H, Ar's), 5.50 (d, 1H, $J_{1'2'} = 4.0$ Hz, H-1'), 5.30 (dd, 1H, $J_{3'4'} = 3.0$ Hz, $J_{4'5'} = 1.0$ Hz, H-4'), 5.15 (dd, 1H, $J_{2'3'} = 10.5$ Hz, H-3'), 4.65 (d, 1H, $J_{12} = 3.5$ Hz, H-1), 4.4-4.8 (m, 7H), 4.15 (m, 1H), 4.05 (dd, 1H, $J_{6a6b} = 11.0$ Hz, $J_{56a} = 6.0$ Hz, H-6a), 3.85 (dd, 1H, $J_{56b} = 7.0$ Hz, H-6b), 3.6-4.0 (m, 5H), 3.50 (dd, 1H, $J_{23} = 9.5$ Hz, H-2), 3.30 (s, 3H, OMe), 1.95-2.1 (3s, 9H, Ac's). Anal. Calc. for C₄₀H₄₈O₁₄ x H₂O: C, 62.33; H, 6.54. Found: C, 61.84; H, 6.54.

21: ¹³C NMR (CDCl₃): δ 170.3-5 (Ac's), 137.3-137.8, 127.6-128.4 (Ar's), 100.7 (C-1'), 97.9 (C-1), 81.4, 78.2, 73.6, 73.2, 70.7, 70.0, 68.8, 68.0, 67.8, 67.0 (10C), 61.5 (C-6'), 55.1 (OMe), 20.5-20.8 (Ac's). ¹H NMR (CDCl₃): δ 7.2-7.4 (m, 10H, Ar's), 5.40 (dd, 1H, $J_{3'4'} = 3.0$ Hz, $J_{4'5'} = 1.0$ Hz, H-4'), 5.35 (dd, 1H, $J_{1'2'} = 3.5$ Hz, H-1'), 5.20 (dd, 1H, $J_{2'3'} = 10.5$ Hz, H-3'), 4.4-4.9 (m, 5H), 4.50 (d, 1H, $J_{12} = 3.5$ Hz), 3.5-4.15 (m, 8H), 3.4 (dd, 1H, $J_{23} = 9.5$ Hz, H-2), 3.3 (s, 3H, OMe), 1.9-2.1 (3s, 9H, Ac's).

Acetylation of **20** (40 mg) with pyridine (0.5 ml) and acetic anhydride (0.2 ml) for 18 h at 25°C, followed by concentration and coconcentration with toluene gave tetraacetate **20a** (48 mg).

¹H NMR (CDCl₃): δ 7.0-7.4 (m, 15H, Ar's), 5.60 (d, 1H, $J_{1'2'} = 3.5$ Hz, H-1'), 5.45 (dd, 1H, $J_{2'3'} = 11.0$ Hz, $J_{3'4'} = 3.0$ Hz, H-3'), 5.35 (dd, 1H, $J_{4'5'} = 1.0$ Hz, H-4'), 5.30 (dd, 1H, H-2'), 4.75 (dt, 1H, $J_{5'6b'} = 7.0$ Hz, $J_{5'6a'} = 6.0$ Hz, H-5'), 4.65 (d, 1H, $J_{12} = 3.5$ Hz, H-1), 4.4-4.7 (m, 6H, Bn's), 4.15 (m, 1H, H-3), 4.05 (dd, 1H, $J_{6a'6b'} = 11.0$ Hz, H-6a'), 3.80 (dd, 1H, H-6b'), 3.6-3.7 (m, 4H, H-4, H-5, H-6a, H-6b), 3.55 (dd, 1H, $J_{23} = 9.5$ Hz, H-2), 3.30 (s, 3H, OMe), 1.9-2.2 (4s, 12H, Ac's).

Acetylation of **21** (47 mg) in pyridine (0.5 ml) and acetic anhydride (0.2 ml) for 18 h at 25°C followed by concentration and coconcentration with toluene gave pentaacetate **21a** (45 mg). ¹H NMR (CDCl₃): δ 7.1-7.4 (m, 10H, Ar's), 5.35 (dd, 1H, $J_{2'3'} = 11.0$ Hz, $J_{3'4'} = 3.5$ Hz, H-3'), 5.30 (d, 1H, $J_{1'2'} = 3.5$ Hz, H-1'), 5.25 (dd, 1H, $J_{4'5'} = 1.0$ Hz, H-4'), 5.05 (dd, 1H, $J_{45} = 10.0$ Hz, $J_{34} = 9.5$ Hz, H-4), 5.0 (dd, 1H, H-2'), 4.65 (d, 1H, $J_{12} = 3.5$ Hz, H-1), 4.4-4.7 (m, 5H), 4.20 (t, 1H, $J_{23} = 9.5$ Hz, H-3), 3.95 (dd, 1H, $J_{6a'6b'} = 11.0$ Hz, $J_{5'6a'} = 6.0$ Hz, H-6a'), 3.75 (dd, 1H, $J_{5'6b'} = 7.0$ Hz, H-6b'), 3.6-3.7 (m, 2H), 3.55 (dd, 1H, H-2), 3.4 (m, 1H), 3.3 (s, 3H, OMe), 1.9-2.1 (4s, 15H, Ac's). MS (CI, NH₃): m/z 764 (M + NH₄⁺).

Methyl 2,6-di-O-benzyl-4,2'-di-O-dimethylsilylene-3-O-(3,4,6-tri-O-acetyl- α -D-galactopyranosyl)- α -D-glucopyranoside (21b). Reaction of **19** (135 mg, 0.15 mmol), with N-iodosuccinimide (83 mg, 0.37 mmol, 2.5 eq) in MeNO₂ (4 ml) for 1½ h at 100°C as described above gave a solution to which CH₂Cl₂ (50 ml) was added. Washing with NaHCO₃-solution (10 ml, saturated) and Na₂S₂O₃-solution (10 ml, 10%), drying (MgSO₄) and concentration gave a syrup (154 mg). Flash-chromatography in EtOAc-pentane 1:4 followed by 1:2 followed by 1:1 gave first **21b** as a colorless syrup (49 mg, 46%) and as a slower moving product **20** (34 mg, 31%). **21b**: [α]_D²⁰ + 66.4° (c 2.5, CH₂Cl₂). ¹³C NMR (CDCl₃): δ 169.9-170.2 (Ac's), 138.0, 137.6, 132.1, 127.3-129.6 (Ar's), 102.2 (C-1'), 98.3 (C-1), 85.0, 77.4, 73.4, 73.3, 71.4, 70.0, 69.9, 68.7, 67.9, 67.7, 66.6 (C-2 - C6, C-2' - C-5', Bn's), 61.0 (C-6'), 55.0 (OMe), 20.3-20.5 (Ac's), -2.7, -3.0 (Me₂Si).

Methyl 2,6-di-O-benzyl-4,2'-di-O-dimethylsilylene-3-O-(3,4,6-tri-O-acetyl- α -D-glucopyranosyl)- α -D-glucopyranoside (22). A solution of **8** (102 mg, 0.11 mol) and NIS (63 mg, 0.28 mmol, 2.5 eq.) in MeNO₂ (5 ml) was refluxed for 3 h. CH₂Cl₂ (50 ml) was added, and the solution washed with NaHCO₃-solution (10 ml, 10%), dried over MgSO₄ and concentrated to a syrup (107 mg). Flash-chromatography in EtOAc-pentane 1:4 followed by 1:2 followed by 1:1 gave syrupy **22** as faster moving product (19 mg, 22%). As a slower moving product **9** was obtained (25 mg, 29%). **22**: ¹³C NMR (CDCl₃): δ 127.5-128.5 (Ar's), 101.4 (C-1'), 98.3 (C-1), 84.9, 77.6, 73.6, 73.4, 73.0, 71.9, 71.7, 70.3, 68.1, 67.9 (10C), 61.7 (C-6'), 55.2 (OMe), 20.6-20.8 (Ac's), -2.7, -2.9 (SiMe₂). ¹H NMR (CDCl₃): δ 7.2-7.6 (m, 10H, Ar's), 5.35 (t, 1H, J_{3'4'} = J_{4'5'} = 9.5 Hz, H-4'), 5.30 (d, 1H, J_{1'2'} = 4.0 Hz, H-1'), 4.95 (t, 1H, J_{2'3'} = 10.0 Hz, H-3'), 4.75 (d, 1H, J = 12.0 Hz, Bn), 4.60 (d, 1H, J₁₂ = 3.5 Hz, H-1), 4.55 (d, 1H, J = 12.0 Hz, Bn), 4.50 (s, 2H, Bn), 4.25 (dd, 1H, J_{6a'6b'} = 12.5 Hz, J_{5'6'} = 3.5 Hz, H-6a'), 4.10 (m, 1H, H-5'), 3.95 (dd, 1H, H-2'), 3.9 (t, 1H, J₃₄ = J₄₅ = 9.0 Hz, H-4), 3.8 (t, 1H, J₂₃ = 9.5 Hz, H-3), 3.78 (dd, 1H, J_{5'6b'} = 2.0 Hz, H-6'b'), 3.70 (dt, 1H, J_{56a} = J_{56b} = 3.0 Hz, H-5), 3.6 (d, 2H, J = 3.0 Hz, H-6a, H-6b), 3.5 (dd, 1H, H-2), 3.35 (s, 3H, OMe), 2.0 (3s, 9H, Ac's), 0.15, 0.10 (2s, 6H, Me₂Si).

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REFERENCES:

- For some biochemical roles of glycosides see: a) Rademacher, T.W.; Parekh, R.B.; Dwek, R.A. *Ann. Rev. Biochem.* **1988**, *57*, 785-838. b) Hoffman, S.; Edelman, G. *Proc. Natl. Acad. Sci USA* **1983**, *80*, 5762-6. c) Calvo, F.O.; Ryan, R.J. *Biochemistry* **1985**, *24*, 1953-9. d) Keilhaver, G.; Faissner, A.; Schachner, M. *Nature* **1985**, *316*, 728-30. e) Springer, T.A. *Nature* **1990**, *346*, 425-434. f) Reddy, A.V.; MacColl, R.; Maley, F. *Biochemistry* **1990**, *29*, 2482-7.
- For some roles of glycosides in various disorders: a) Kornfeld, S. *J. Clin. Invest* **1986**, *77*, 1-6.b) Humphries, M.J.; Matsumoto, K.; White, S.L.; Olden, K. *Proc. Natl. Acad. Sci. USA* **1986**, *83*, 1752-6. c) Dennis, J.W.; Laferte, S.; Waghorne, C.; Breitman, M.L.; Kerbel, R.S. *Science* **1987**, *236*, 582-5. d) Fukuda, M.N.; Dell, A.; Scartezzin, P. *J. Biol. Chem.* **1987**, *262*, 7195-7206. e) Montefiori, D.C.; Robinson, W.E.Jr.; Mitchell, W.M. *Proc. Natl. Acad. Sci. USA* **1988**, *85*, 9248-52.

3. For some reviews: a) Paulsen, H. *Angew. Chem.* 1982, *94*, 184-201 (*Int. ed.* 1982, *22*, 155-173). b) Paulsen, H. *Chem. Soc. Rev.* 1984, *13*, 15-45. c) Schmidt, R.R. *Angew. Chem.* 1986, *98*, 213-236 (*Int. ed.* 1986, *25*, 212-235). d) Binkley, R.W. "Modern Carbohydrate Chemistry" (Chapter 14). Marcel Dekker: New York 1988, 297-322. e) Bochkov, A.F.; Zaikov, G.E. "Chemistry of the O-glycosidic bond" Pergamon Press Oxford 1979. f) Fügedi, P.; Garegg, P.J.; Lönn, H.; Norberg, T. *Glycoconjugate J.* 1987, *4*, 97-108. g) Banoub, J.; Boullanger, P.; Lafont, D. *Chem. Rev.* 1992, *92*, 1167-1195. h) Garegg, P.J. *Acc Chem. Res.* 1992, *25*, 575-580.
4. a) Veeneman, G.H.; Boom, J.H. van *Tetrahedron Lett.* 1990, *31*, 275-8. b) Veeneman, G.H.; Leenwen, S.H. van; Boom, J.H. van *Tetrahedron Lett.* 1990, *31*, 1331-4. c) Konradsson, P.; Udodong, V.E.; Fraser-Reid *Tetrahedron Lett.* 1990, *31*, 4313-6. d) Fügedi, P.; Garegg, P.J. *Carbohydr. Res.* 1986, *149*, C9-C12. e) Schmidt, R.R.; Michel, J. *Angew. Chem.* 1980, *19*, 763-4.
5. Barresi, F.; Hindsgaul, O. *J. Am. Chem. Soc.* 1991, *113*, 9376-7.
6. Barresi, F.; Hindsgaul, O. *Synlett* 1992, 759-761.
7. Stork, G.; Kim, G. *J. Am. Chem. Soc.* 1992, *114*, 1087-8.
8. Bols, M. *J. Chem. Soc., Chem. Commun.*, 1992, 913-4.
9. Bols, M. *Acta Chem. Scand.* (in press).
10. Bols, M. *J. Chem. Soc., Chem. Commun.*, (in press).
11. Koto, S.; Morishima, N.; Owa, M.; Zen, S. *Carbohydr. Res.* 1984, *130*, 73-83.
12. Brady, R.F. *Carbohydr. Res.* 1970, *15*, 35-40.
13. Bolliger, H.R.; Prins, D.A. *Helv. Chim. Acta*, 1946, *29*, 1116-20.
14. Collins, P.M.; Overend, W.G.; Rayner, B.A. *Carbohydr. Res.* 1973, *31*, 1-16.
15. Boren, H.B.; Ekborg, G.; Eklind, K.; Garegg, P.J.; Pilotti, Å.; Swahn, C.-G. *Acta Chem. Scand.* 1973, *27*, 2639-2644.
16. Zhang, Y.-M.; Mallet, J.-M.; Sinay, P. *Carbohydr. Res.* 1992, *236*, 73-88.